

KINETIC MODELLING OF ETHANOL PRODUCTION FROM OIL PALM TRUNK SAP DURING FERMENTATION

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STUDENT'S DECLARATION

I hereby declare that the work in this thesis is based on my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously or concurrently submitted for any other degree at Universiti Malaysia Pahang or any other institutions.

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SAP DURING FERMENTATION

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ABSTRAK

Penyimpanan bahan api fosil di seluruh dunia yang terhad dan impaknya yang buruk terhadap alam sekitar membawa kepada penyelidikan terkini ke arah penggunaan biomas dalam penghasilan biofuel. Malaysia kaya dengan sumber biomas. Batang kelapa sawit (OPT) adalah sumber biomas untuk menghasilkan bioethanol. Fermentasi adalah proses terpenting dalam penukaran biomas kepada etanol. Model kinetik yang sesuai mampu meningkatkan kecekapan dan proses pengoptimuman penapaian etanol menggunakan sap OPT. Kaedah teoritikal lebih efisien dan memerlukan kos pelaburan yang rendah, tetapi kaedah ini sukar untuk disahkan. Beberapa model kinetik telah dicadangkan tetapi tiada model yang mengambil kira faktor-faktor penting seperti batasan substrat, perencat substrat, penghambatan produk, dan kematian sel secara serentak pada suhu berbeza untuk menghasilkan etanol dari penapaian sap OPT. Kami memanjangkan dan memperbaiki model matematik terkini untuk meneroka kesan suhu, kepekatan sel terawal dan kadar kematian sel pada proses penapaian. Beberapa parameter kinetik digunakan untuk menggambarkan fenomena ini. Satu set persamaan pembezaan biasa digunakan untuk memodelkan profil gula, sel dan etanol untuk penapaian sap OPT dan persamaan telah diselesaikan oleh kaedah Runge-Kutta untuk ke-4. Terdapat dua set hasil simulasi yang dibentangkan dalam kajian ini untuk Model I dan II. Model I adalah model mudah yang memanjangkan model Oliviera, di mana kami mengkaji kesan kadar kematian sel. Model II lebih komprehensif dan lebih baik daripada Model I, kerana ia mempunyai hubungan Leudeking-Piret, model Phisalaphong dan juga Model I. Sesetengah ciri-ciri penting dikenalpasti kedua-dua model. Apabila suhu meningkat, kadar pertumbuhan sel khusus maksimum menurun bagi kedua-dua model. Dari hasilnya, Suhu yang sesuai untuk pengeluaran etanol dari penapaian sap OPT ialah 30°C. Kadar penggunaan gula dan pengeluaran etanol sepanjang proses penapaian bergantung pada kepekatan sel awal. Dengan kepekatan sel awal yang rendah, kadar penukaran meningkat secara beransur-ansur tetapi untuk kepekatan sel awal yang tinggi, penukaran gula ke etanol meningkat dengan ketara dan berkurangan selepas tempoh yang singkat disebabkan oleh akses etanol, yang mungkin menghalang pertumbuhan sel. Pertimbangan gabungan batasan dan perencatan substrat, penghambatan produk dan kadar kematian sel meningkatkan ketepatan model I dengan cara rRMSE. Pemerhatian serupa ditemui untuk model II apabila faktor-faktor yang dipertimbangkan adalah had dan penyekatan substrat, pertumbuhan dan pembentukan produk yang berkaitan dengan pertumbuhan, penghambatan produk dan kematian sel. Pendekatan ini membolehkan kita memperoleh keupayaan ramalan yang lebih baik dengan itu meningkatkan pemahaman kita terhadap model matematik penapaian sap OPT.

ABSTRACT

The worldwide limited storage of fossil fuel and its bad impact on environment lead to the recent research towards biomass for biofuel. Malaysia is rich with plenty of biomass resources. Oil palm trunk (OPT) is a promising biomass source for bioethanol production. Fermentation is an essential process of biomass to ethanol conversion. An appropriate kinetic model will be a powerful tool to increase the efficiency and process optimization for ethanol fermentation using the OPT sap. The theoretical methods are more efficient and require low investment, but it is challenging to validate. A number of kinetic models have been proposed but none of these models observed the effect of most essential factors such as substrate limitation, substrate inhibition, product inhibition, and cell death simultaneously on temperature to produce ethanol from the OPT sap fermentation. We extended and improved the current mathematical model to explore the effect of temperature, initial cell concentration and cell death rate on the fermentation process. Several kinetic parameters were used to describe this phenomenon. A set of ordinary differential equations were used to modelled the profiles of sugar, cell and ethanol for the fermentation of OPT sap and the equations were solved by the 4th order Runge-Kutta method. There are two sets of simulation results presented in this study for Model I and II. Model I is a simple model which extends Oliviera's model, where we studied the effect of cell death rate. Model II is more comprehensive and better than Model I, because it consists Leudeking-Piret relationship, Phisalaphong model and also Model I. Some significant characteristics are apprehended both of the models. As the temperature increased, the maximum specific cell growth rate decreased for both of the models. From the results, the suitable temperature for ethanol production from the OPT sap fermentation is 30°C. The rate of sugar utilisation and ethanol production throughout fermentation process depend on the initial cell concentration. With the low initial cell concentration, the conversion rate was increased gradually but for the high initial cell concentration, sugar conversion to ethanol was augmented sharply and depleted after the short duration due to access of the ethanol, which might inhibit the cell growth. The combined consideration of the substrate limitation and inhibition, growth and non-growth associated product formation, product inhibition and cell death rate increased the accuracy of the Model II by means of rRMSE. This approach has enabled us to obtained a better predictive capabilities hence increasing our understanding of the mathematical model of the OPT sap fermentation.

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LIST OF SYMBOLS

a	growth associated specific productivity coefficient
b	Non-growth associated specific productivity coefficient
c	Coefficient of cell maintenance
$f(x_i)$	Calculated data
K_{CM}	Maintenance constant
K_d	Cell death rate
K_i	Substrate inhibition coefficient/ Inhibition parameter for sugar
K_S	Saturation constant of substrate/ Substrate inhibition constant
$K_{in,S}$	Substrate inhibition constant
$K_{li,S}$	Substrate limitation constant
K_{li,O_2}	Dissolved oxygen limitation constant
K_P	Product inhibition constant
K_{SP}	Saturation constant for ethanol production
K_{SS}	Substrate inhibition term for cell growth
K_{SSP}	Substrate inhibition term for ethanol production
n	Ethanol toxic power
O_2	Dissolved oxygen concentration
P	Ethanol production
P_m	Ethanol inhibition term for cell growth
P'_M	Ethanol inhibition term for ethanol production
P^*	Critical concentration of product
r	Reaction rate
$Y'_{x/s}, Y_{x/s}$	Yield coefficient for the cell on substrate
$Y'_{p/s}, Y_{p/s}$	Yield coefficient for the ethanol on substrate
y_i	Experimental data
α	Model parameter
μ	Specific growth rate

$\hat{\mu}$	Maximum specific growth rate
ν	Specific production rate

REFERENCES

- Aiba, S. and Shoda, M. (1969) Reassessment of product inhibition in alcohol fermentation. *Journal of Fermentation Technology*, 47(12), 790.
- Alauddin, Z.A.B.Z., Lahijani, P., Mohammadi, M. and Mohamed, A.R. (2010) Gasification of lignocellulosic biomass in fluidized beds for renewable energy development: A review. *Renewable and Sustainable Energy Reviews*, 14(9), 2852-2862.
- Badger, P. (2002) Ethanol from cellulose: a general review. *Trends in new crops and new uses*, 1, 17-21.
- Bansal, P., Hall, M., Realff, M.J., Lee, J.H. and Bommarius, A.S. (2009) Modeling cellulase kinetics on lignocellulosic substrates. *Biotechnology advances*, 27(6), 833-848.
- Beale, C.M., Lennon, J.J., Yearsley, J.M., Brewer, M.J. and Elston, D.A. (2010) Regression analysis of spatial data. *Ecology Letters*, 13(2), 246-264.
- Birol, G., Doruker, P., Kirdar, B., Önsan, Z.İ. and Ülgen, K. (1998) Mathematical description of ethanol fermentation by immobilised *Saccharomyces cerevisiae*. *Process Biochemistry*, 33(7), 763-771.
- Bosse, T. and Griewank, A. (2014) Optimal control of beer fermentation processes with Lipschitz-constraint on the control. *Journal of the Institute of Brewing*, 120(4), 444-458.
- Clark, D.S. and Blanch, H.W. (1996) *Biochemical Engineering*, Marcel Dekker, Inc, USA.
- Costa, R.S., Hartmann, A. and Vinga, S. (2016) Kinetic modeling of cell metabolism for microbial production. *Journal of Biotechnology*, 219, 126-141.
- Esfahanian, M., Rad, A.S., Khoshhal, S., Najafpour, G. and Asghari, B. (2016) Mathematical modeling of continuous ethanol fermentation in a membrane bioreactor by pervaporation compared to conventional system: Genetic algorithm. *Bioresource Technology*, 212, 62-71.
- Fan, S., Chen, S., Tang, X., Xiao, Z., Deng, Q., Yao, P., Sun, Z., Zhang, Y. and Chen, C. (2015) Kinetic model of continuous ethanol fermentation in closed-circulating process with pervaporation membrane bioreactor by *Saccharomyces cerevisiae*. *Bioresource Technology*, 177, 169-175.
- Garnier, A. and Gaillet, B. (2015) Analytical solution of Luedeking–Piret equation for a batch fermentation obeying Monod growth kinetics. *Biotechnology and Bioengineering*, 112(12), 2468-2474.
- Ghaly, A. and El-Taweel, A. (1994) Kinetics of batch production of ethanol from cheese whey. *Biomass and Bioenergy*, 6(6), 465-478.

- Ghose, T. and Tyagi, R. (1979) Rapid ethanol fermentation of cellulose hydrolysate. II. Product and substrate inhibition and optimization of fermentor design. *Biotechnology and Bioengineering*, 21(8), 1401-1420.
- Griggs, A.J., Stickel, J.J. and Lischeske, J.J. (2012a) A mechanistic model for enzymatic saccharification of cellulose using continuous distribution kinetics I: depolymerization by EGI and CBHI. *Biotechnology and Bioengineering*, 109(3), 665-675.
- Griggs, A.J., Stickel, J.J. and Lischeske, J.J. (2012b) A mechanistic model for enzymatic saccharification of cellulose using continuous distribution kinetics II: cooperative enzyme action, solution kinetics, and product inhibition. *Biotechnology and Bioengineering*, 109(3), 676-685.
- Guidini, C.Z., Marquez, L.D.S., de Almeida Silva, H., de Resende, M.M., Cardoso, V.L. and Ribeiro, E.J. (2014) Alcoholic fermentation with flocculant *Saccharomyces cerevisiae* in fed-batch process. *Applied Biochemistry and Biotechnology*, 172(3), 1623-1638.
- Halim, N.B.A. (2016) Optimization of bioethanol production from oil palm trunk sap, Universiti Malaysia Pahang.
- Himmel, M.E., Ding, S.-Y., Johnson, D.K., Adney, W.S., Nimlos, M.R., Brady, J.W. and Foust, T.D. (2007) Biomass recalcitrance: engineering plants and enzymes for biofuels production. *Science*, 315(5813), 804-807.
- Hinshelwood, C.N. (1946) The chemical kinetics of the bacterial cell, Oxford: The Clarendon Press, UK.
- Hoppe, G.K. and Hansford, G.S. (1982) Ethanol inhibition of continuous anaerobic yeast growth. *Biotechnology Letters*, 4(1), 39-44.
- Hossain, N. and Jalil, R. (2015) Sugar and bioethanol production from oil palm trunk (OPT). *Asia Pacific Journal of Energy and Environment*, 2(2), 81-84.
- Jamil, N.M. and Wang, Q. (2016) The Nondimensionalization of Equations Describing Enzymatic Cellulose Hydrolysis. *World Applied Sciences Journal*, 34(2), 158-163.
- Jin, H., Liu, R. and He, Y. (2012) Kinetics of batch fermentations for ethanol production with immobilized *Saccharomyces cerevisiae* growing on sweet sorghum stalk juice. *Procedia Environmental Sciences*, 12, 137-145.
- Kelkar, S. and Dolan, K. (2012) Modeling the effects of initial nitrogen content and temperature on fermentation kinetics of hard cider. *Journal of Food Engineering*, 109(3), 588-596.
- Khalifa, N. (2011) Empirical and kinetic models for the determination of pharmaceutical product stability.

- Limayem, A. and Ricke, S.C. (2012) Lignocellulosic biomass for bioethanol production: current perspectives, potential issues and future prospects. *Progress in Energy and Combustion Science*, 38(4), 449-467.
- Liu, J.-Z., Weng, L.-P., Zhang, Q.-L., Xu, H. and Ji, L.-N. (2003) A mathematical model for gluconic acid fermentation by *Aspergillus niger*. *Biochemical engineering journal*, 14(2), 137-141.
- Liu, Z. and Li, X. (2014) The kinetics of ethanol fermentation based on adsorption processes. *Kemija u Industriji*, 63(7-8).
- Luedeking, R. and Piret, E.L. (2000) A kinetic study of the lactic acid fermentation. Batch process at controlled pH. *Biotechnology and Bioengineering*, 67(6), 636-644.
- Matsuoka, Y. and Shimizu, K. (2015) Current status and future perspectives of kinetic modeling for the cell metabolism with incorporation of the metabolic regulation mechanism. *Bioresources and Bioprocessing*, 2(1), 1-19.
- Matsushika, A. and Sawayama, S. (2010) Effect of initial cell concentration on ethanol production by flocculent *Saccharomyces cerevisiae* with xylose-fermenting ability. *Applied Biochemistry and Biotechnology*, 162(7), 1952-1960.
- Mohamad, N.L., Kamal, S.M.M., Mokhtar, M.N., Husain, S.A. and Abdullah, N. (2016) Dynamic mathematical modelling of reaction kinetics for xylitol fermentation using *Candida tropicalis*. *Biochemical engineering journal*, 111, 10-17.
- Monod, J. (1949a) The growth of bacterial cultures. *A. Rev. Microbiol*, 3, 371-394.
- Monod, J. (1949b) The growth of bacterial cultures. *Annual Reviews in Microbiology*, 3(1), 371-394.
- Oliveira, S.C., Oliveira, R.C., Tacin, M.V. and Gattás, E.A. (2016) Kinetic modeling and optimization of a batch ethanol fermentation process. *Journal of Bioprocessing & Biotechniques*, 6, 266.
- Ona, I., Agogo, H. and Iorungwa, M. (2019) Production of Bioethanol from Cassava using *Zymomonas mobilis*: Effect of Temperature and Substrate concentration. *NIGERIAN ANNALS OF PURE AND APPLIED SCIENCES*, 1(1), 153-160.
- Papagianni, M., Boonpooh, Y., Matthey, M. and Kristiansen, B. (2007) Substrate inhibition kinetics of *Saccharomyces cerevisiae* in fed-batch cultures operated at constant glucose and maltose concentration levels. *Journal of industrial microbiology & biotechnology*, 34(4), 301-309.
- Phisalaphong, M., Srirattana, N. and Tanthapanichakoon, W. (2006) Mathematical modeling to investigate temperature effect on kinetic parameters of ethanol fermentation. *Biochemical engineering journal*, 28(1), 36-43.
- Rivera, E.C., Da Costa, A.C., Lunelli, B.H., Maciel, M.R.W. and Maciel Filho, R. (2008) Kinetic modeling and parameter estimation in a tower bioreactor for

- bioethanol production. *Applied biochemistry and biotechnology*, 148(1-3), 163-173.
- Rivera, E.C., Yamakawa, C.K., Saad, M.B., Atala, D.I., Ambrosio, W.B., Bonomi, A., Junior, J.N. and Rossell, C.E. (2016) Effect of temperature on sugarcane ethanol fermentation: Kinetic modeling and validation under very-high-gravity fermentation conditions. *Biochemical engineering journal*.
- Salgado, J.M., Converti, A. and Domínguez, J.M. (2012) D-Xylitol, pp. 161-191, Springer.
- Sampaio, F.C., Mantovani, H.C., Passos, F.J.V., de Moraes, C.A., Converti, A. and Passos, F.M.L. (2005) Bioconversion of d-xylose to xylitol by *Debaryomyces hansenii* UFV-170: Product formation versus growth. *Process Biochemistry*, 40(11), 3600-3606.
- Samsudin, M.D.M. and Don, M.M. (2015) Assessment of bioethanol yield by *S. cerevisiae* grown on oil palm residues: Monte Carlo simulation and sensitivity analysis. *Bioresource Technology*, 175, 417-423.
- Silva, C.J., Mussatto, S.I. and Roberto, I.C. (2006) Study of xylitol production by *Candida guilliermondii* on a bench bioreactor. *Journal of Food Engineering*, 75(1), 115-119.
- Silva, S.S., Roberto, I.C., Felipe, M.G. and Mancilha, I.M. (1996) Batch fermentation of xylose for xylitol production in stirred tank bioreactor. *Process Biochemistry*, 31(6), 549-553.
- Srimachai, T., Nuithitikul, K., Sompong, O., Kongjan, P. and Panpong, K. (2015) Optimization and kinetic modeling of ethanol production from oil palm frond juice in batch fermentation. *Energy Procedia*, 79, 111-118.
- Talib, A.T., Mokhtar, M.N., Baharuddin, A.S. and Sulaiman, A. (2014) Effects of aeration rate on degradation process of oil palm empty fruit bunch with kinetic-dynamic modeling. *Bioresource Technology*, 169, 428-438.
- Tian, Y.-C., Zhang, T., Yao, H. and Tadé, M.O. (2014) Computation of mathematical models for complex industrial processes, World Scientific.
- Vázquez, J.A. and Murado, M.A. (2008) Unstructured mathematical model for biomass, lactic acid and bacteriocin production by lactic acid bacteria in batch fermentation. *Journal of Chemical Technology and Biotechnology*, 83(1), 91-96.
- Wang, F.-S. and Sheu, J.-W. (2000) Multiobjective parameter estimation problems of fermentation processes using a high ethanol tolerance yeast. *Chemical Engineering Science*, 55(18), 3685-3695.
- Wang, L., Agyemang, S.A., Amini, H. and Shahbazi, A. (2015) Mathematical modeling of production and biorefinery of energy crops. *Renewable and Sustainable Energy Reviews*, 43, 530-544.

- Wang, Y. and Liu, S. (2014) Kinetic modeling of ethanol batch fermentation by *Escherichia coli* FBWHR using hot-water sugar maple wood extract hydrolyzate as substrate. *Energies*, 7(12), 8411-8426.
- You, L. and Baharin, B. (2006) Effects of enzymatic hydrolysis on crude palm olein by lipase from *Candida rugosa*. *Journal of Food Lipids*, 13(1), 73-87.
- Zhang, F. (2011) Development of an optimization model for biofuel facility size and location and a simulation model for design of a biofuel supply chain. *PhD Thesis*.
- Zheng, L. and Zhang, X. (2017) Modeling and Analysis of Modern Fluid Problems, Academic Press.
- Zhu, C., Fang, B. and Wang, S. (2016) Effects of culture conditions on the kinetic behavior of 1, 3-propanediol fermentation by *Clostridium butyricum* with a kinetic model. *Bioresource Technology*, 212, 130-137.